

# Nitrogen Incorporation in the Biosynthetic Pathway of the Nitrogen-containing Polyketide, Pamamycin in *Streptomyces alboniger*

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**Abstract** The biosynthetic pathway of pamamycin (**1**), a nitrogen-containing polyketide, was investigated using blocked mutants of *Streptomyces alboniger*. Hydroxy acids K (**3**), L (**4**) and S (**5**) were found in cultured materials of blocked mutants and the wild type strain, but no PM-ketone (**2**) was detected. Hydroxy acids **3**, **4**, **5** and de-*N*-methylhydroxy acid L (**7**) were converted into **1**, but **2** nor de-*N*-methylpamamycin (**6**) were not. We also confirmed that **3** and **7** were converted into **4**. These results showed that an amino group was introduced into the carbonyl group of **3** by transamination, and subsequent *N*-methylation led to **4** in the pamamycin biosynthetic pathway. Quantitative analyses of hydroxy acid intermediates **3**, **4**, and **5**, and pamamycin (**1**) suggested that transamination was the rate-determining step in pamamycin biosynthesis.

**Keywords** pamamycin, biosynthesis, transamination, polyketide

## Introduction

Pamamycin-607 (**1**; MW 607; R<sub>1</sub>, R<sub>2</sub>, R<sub>3</sub>, R<sub>4</sub>=CH<sub>3</sub>, R<sub>5</sub>=H) (Fig. 1) has been isolated from *Streptomyces alboniger* as an autoregulator inducing aerial mycelium formation in an

aerial mycelium-less mutant [1–3]. In previous reports, we isolated 14 pamamycin homologues with different alkyl substituents at R<sub>1</sub>–R<sub>5</sub> with MW ranging from 593 to 649, prepared some degradation products of pamamycin-607, and discussed structure–activity relationships [4–6]. We also showed that exogenous pamamycin-607 restored or stimulated aerial mycelium formation in many *Streptomyces* species, indicating that pamamycin-607 acts in a common regulatory system for aerial mycelium formation in *Streptomyces* spp. [7].

The structure of pamamycin consists of a unique sixteen-membered macrodiolide with a dimethylamino group-bearing side chain. We recently showed using feeding experiments with <sup>13</sup>C- and <sup>15</sup>N-labeled compounds that the carbon skeleton of pamamycin-607 was derived from 6 acetate, 4 propionate and 3 succinate units *via* a polyketide pathway, and a nitrogen atom was incorporated from an  $\alpha$ -amino group of an amino acid into a carbonyl group by transamination reaction [8].

The macrodiolide structure of pamamycin suggested that the carbon skeleton of **1** was assembled from two hydroxy acids K (**3**) and S (**5**). It is still unknown when the nitrogen atom is introduced into the carbon skeleton. We are interested in investigating nitrogen incorporation during biosynthesis of pamamycin because the dimethylamino group of pamamycin-607 is indispensable for aerial

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mycelium-inducing activity [6] and nitrogen-containing polyketides are rare.

We suggest two pathways for the biosynthesis of **1** from two hydroxy acids **3** and **5** (Fig. 1). One pathway involves the formation of a keto-macrodiolide intermediate (PM-ketone, **2**) by condensation of **3** and **5**, then the nitrogen atom is introduced into the carbon skeleton by transamination, and *N*-methylation leads to **1**. Isolation of de-*N*-methylpamamycins (**6**) from cultured materials of *S. alboniger* [9, 10] supported this hypothesis. The other pathway would involve acceptance of a nitrogen atom by the larger hydroxy acid **3**, and then *N*-methylation occurs to produce hydroxy acid L (**4**) before formation of the macrodiolide ring.

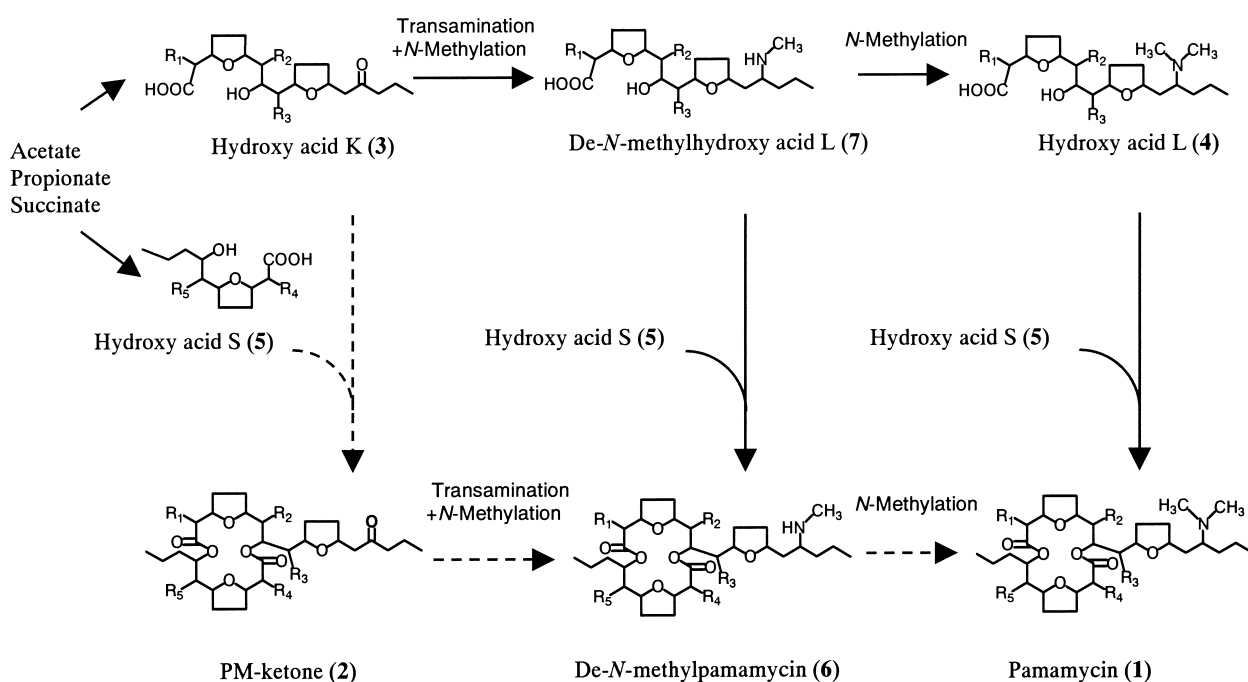
In this paper, we describe transamination and condensation steps in pamamycin biosynthesis by investigating biosynthetic intermediates of pamamycin in cultured materials of blocked mutants of *S. alboniger*, and report bioconversion experiments with these compounds. The *N*-methylation step was also investigated using bioconversion experiments of de-*N*-methylpamamycin (**6**) and de-*N*-methylhydroxy acid L (**7**). The rate-determining step of pamamycin production was discussed by quantitative analyses of biosynthetic intermediates and pamamycin (**1**) in cultured materials of the wild type strain.

## Results

### Isolation of Blocked Mutants

Mutants producing no or little pamamycin were screened for two reasons; strains in which mutation occur in the later step of the biosynthetic pathway would accumulate intermediates, and strains in which the earlier biosynthetic step is disturbed would be used for conversion experiments of possible intermediates into pamamycin.

From 40,000 colonies obtained by *N*-methyl-*N'*-nitro-*N*-nitrosoguanidine (NTG) mutation, 400 colonies which formed no or little aerial mycelia were selected. These mutants were tested for their responses to pamamycin-607, and 20 strains whose aerial mycelium formation was restored or stimulated by pamamycin-607 were further selected (Fig. 2). Production of pamamycin by these 20 strains was then determined by liquid chromatography-electrospray ionization mass spectrometry (LC-ESI-MS) analysis. Among them, strains 188 and 202 showed lower pamamycin production than that of the wild type strain; production of total pamamycin and pamamycin-607 were 5.7 and 1.0  $\mu\text{g}/\text{ml}$  in strain 188, and 19 and 1.0  $\mu\text{g}/\text{ml}$  in strain 202, respectively; whereas the wild type strain produced 40  $\mu\text{g}/\text{ml}$  total pamamycin and 6.0  $\mu\text{g}/\text{ml}$  pamamycin-607.



**Fig. 1** Structure and proposed biosynthetic pathway of pamamycin.

### Biosynthetic Intermediates Produced by Mutants and the Wild Type Strain

Presence of possible intermediates of pamamycin, PM-ketone (**2**), hydroxy acids K (**3**), L (**4**) and S (**5**), were investigated in the cultured materials of two mutants, strains 188 and 202, and amounts of these compounds were compared with those of the wild type strain (Fig. 3). In



**Fig. 2** Response of aerial mycelium-deficient mutants to pamamycin.

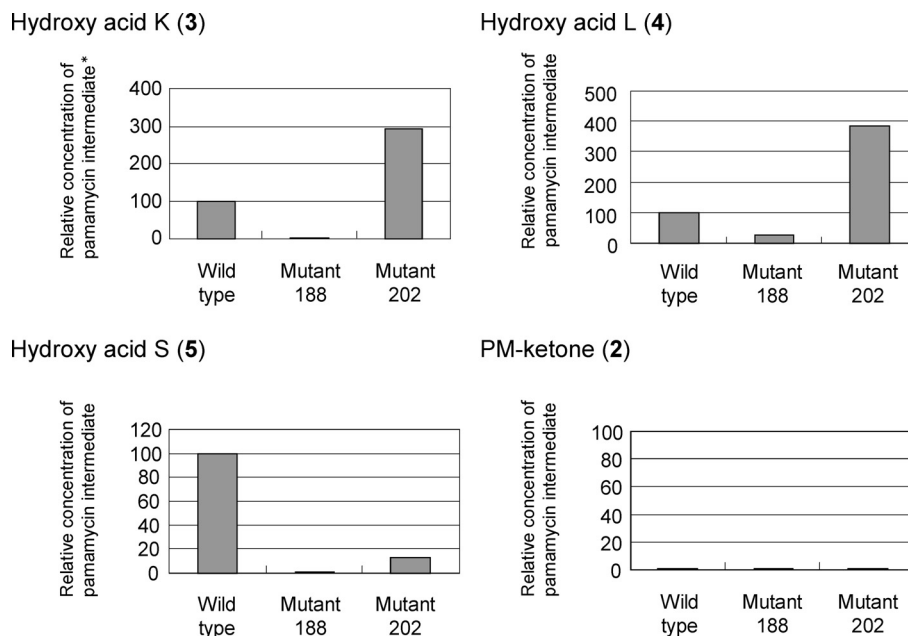
Paper disk containing pamamycin-607 was placed in the center of agar medium and 4 mutants were streaked radially around the disk. The upper mutant restored the formation of white aerial mycelia in response to pamamycin-607 but other 3 mutants remained in bald.

strain 188, **3** and **5** were not detected, and **4** was produced at one-fourth the amount produced by the wild type strain. In strain 202, **5** decreased to one-tenth of wild type strain, but **3** and **4** increased 2.9 and 3.8 folds more than the wild type strain. On the other hand, no **2** was detected in both blocked mutants and the wild type strain (Fig. 3).

These results suggested that **1** was not synthesized *via* **2** but by condensation of **4** and **5**, and the nitrogen atom of **1** was introduced to the carbonyl group of **3** before condensation with **5**. These results also suggested that mutation in strain 188 occurred during the earlier steps of the biosynthetic pathway that were common to hydroxy acids **4** and **5**; and in strain 202, mutation occurred during biosynthesis of hydroxy acid **5**.

### Bioconversion of Hydroxy Acids (**3**, **4**, **5**) and PM-ketone (**2**) to Pamamycin (**1**)

Bioconversion experiment of **5** to **1** was performed with strain 202. Non-labeled and  $^{13}\text{C}$ -labeled **5** were prepared by alkaline hydrolysis of non-labeled and  $^{13}\text{C}$ -labeled pamamycin (**1**), respectively.  $^{13}\text{C}$ -Labeled **1** was obtained by a feeding experiment with  $[2-^{13}\text{C}]$  acetate as described in a previous paper [8]. Hydroxy acid **5** was used as a mixture of homologues, and incubated for 60 hours. Pamamycin fraction obtained from mycelia was analyzed using LC-ESI-MS, and intensities of the molecular ion ( $m/z$  608) and its isotope peaks of pamamycin-607 were



**Fig. 3** Analysis of intermediates accumulated in blocked mutants during pamamycin biosynthesis.

\* Production of intermediates by the wild type strain were defined as 100.

Detection limits were 18, 2, 2, and 0.5 ng for hydroxy acids K, L, S, and PM-ketone, respectively.

measured.

In the feeding experiment with  $^{13}\text{C}$ -labeled **5**, ratios of isotope peaks ( $m/z$  610 (M+H+2), 611 (M+H+3) and 612 (M+H+4)) in the molecular ion cluster were higher than those with administration of non-labeled **5** (Table 1). These results showed that hydroxy acid **5** was incorporated into **1**.

Bioconversions of **3**, **4** and **2** to **1** were performed with strain 188 in a similar manner as in the feeding experiment with **5** in strain 202. Hydroxy acids **3** and **4** were prepared by alkaline hydrolysis of **2** and **1**, respectively, hence each contained comparable amounts of **5**. PM-ketone (**2**) was derived from **1** by permanganate oxidation [11].

Addition of  $^{13}\text{C}$ -labeled **3** or **4** containing **5** increased intensities of isotope peaks with  $m/z$  610 (M+H+2) to 614 (M+H+6) in the molecular ion cluster as compared with those of non-labeled precursors, respectively (Table 1). These results showed that hydroxy acids **3** and **4** were incorporated into **1** since intensity of isotope peaks with larger  $m/z$  values increased compared with results of sole administration of  $^{13}\text{C}$ -labeled **5**. In the feeding experiment with **3**+**5**, compound **2** was not detected in cultured materials.

Feeding experiments with  $^{13}\text{C}$ -labeled and non-labeled **2** showed no differences in intensity for each isotope peak compared with that of total molecular ion peaks (Table 1), although  $^{13}\text{C}$ -labeled **2** was recovered from the mycelial extract (data not shown), suggesting that **2** was

incorporated into mycelia, but was not converted into **1**.

#### Bioconversion of De-*N*-methylpamamycin (**6**) and De-*N*-methylhydroxy acid L (**7**) to Pamamycin (**1**)

To confirm that a nitrogen atom was introduced into **3**, feeding experiments with de-*N*-methylpamamycin (**6**) and de-*N*-methylhydroxy acid L (**7**) were performed in strain 188. De-*N*-methylpamamycin **6** used for the experiment was purified from cultured materials of *S. alboniger* as reported previously [9, 10], and a mixture of **7** and **5** was prepared by alkaline hydrolysis of **6**.

Feeding experiments with  $^{13}\text{C}$ -labeled and non-labeled **6** showed no differences in relative intensity of peaks in the molecular ion cluster (Table 1), although  $^{13}\text{C}$ -labeled **6** was detected in the mycelial extract (data not shown), suggesting that **6** was transported into mycelia but was not converted into **1** as was the case with **2**.

Addition of  $^{13}\text{C}$ -labeled **7** with **5** increased intensities of isotope peaks with  $m/z$  610 (M+H+2) to 612 (M+H+4) in the molecular ion cluster compared with those with non-labeled precursors (Table 1). These results showed that **7** was incorporated into **1**.  $^{13}\text{C}$ -labeled **7** with **5** also increased intensities of isotope peaks of de-*N*-methylpamamycin (**6**) in cultured materials (data not shown).

#### Bioconversion of Hydroxy Acid K (**3**) and De-*N*-methylhydroxy Acid L (**7**) to Hydroxy Acid L (**4**)

In feeding experiments with **3** and **5** or **7** and **5**,

**Table 1** Effects of non-labeled and  $^{13}\text{C}$ -labeled precursors on relative intensity of molecular ion peak and isotope peaks of pamamycin-607

Precursors	Strains	Ratio of molecular ion peak and isotope peaks (%) in the molecular ion cluster <sup>a</sup>							
		608	609	610	$m/z$		613	614	
					611	612			
<b>5</b>	Non-labeled	72	24	4	— <sup>b</sup>	—	—	—	
	$^{13}\text{C}$ -labeled	65	24	7	2	1	—	—	
<b>3</b> + <b>5</b>	Non-labeled	70	25	5	—	—	—	—	
	$^{13}\text{C}$ -labeled	54	25	11	5	2	2	1	
<b>4</b> + <b>5</b>	Non-labeled	69	26	5	—	—	—	—	
	$^{13}\text{C}$ -labeled	52	26	11	5	3	2	1	
<b>2</b>	Non-labeled	71	24	5	—	—	—	—	
	$^{13}\text{C}$ -labeled	70	25	5	—	—	—	—	
<b>6</b>	Non-labeled	70	26	4	—	—	—	—	
	$^{13}\text{C}$ -labeled	71	24	5	—	—	—	—	
<b>7</b> + <b>5</b>	Non-labeled	71	25	4	—	—	—	—	
	$^{13}\text{C}$ -labeled	60	27	9	3	1	—	—	

<sup>a</sup> Sum of peak intensity is 100%. <sup>b</sup> <1%.

**Table 2** Effects of  $^{13}\text{C}$ -labeled precursors on relative intensity of molecular ion peak and isotope peaks of hydroxy acid L with MW 413

Precursors	Strains	Ratio of molecular ion peak and isotope peaks (%) in the molecular ion cluster <sup>a</sup>					
		414	415	<i>m/z</i> 416	417	418	
<b>3+5</b>	Non-labeled	Mutant 188	84	14	2	— <sup>b</sup>	—
	$^{13}\text{C}$ -labeled		70	15	8	7	—
<b>7+5</b>	Non-labeled	Mutant 188	84	14	2	—	—
	$^{13}\text{C}$ -labeled		76	18	4	2	—

<sup>a</sup> Sum of peak intensity is 100%. <sup>b</sup> <1%.

**Table 3** Production of pamamycin and its intermediates by wild type strains of *S. alboniger*

Compound	Production of pamamycin and its intermediates	
	$\mu\text{g/ml}$	$\text{nmol/ml}$
Hydroxy acid K ( <b>3</b> )	0.62	1.62
Hydroxy acid L ( <b>4</b> )	0.27	0.66
Hydroxy acid S ( <b>5</b> )	0.63	2.74
Pamamycin ( <b>1</b> )	40.0	65.9

incorporation of **3** or **7** into hydroxy acid L (**4**) fraction was also analyzed together with the pamamycin fraction.

Addition of  $^{13}\text{C}$ -labeled **3** or **7** increased intensities of isotope peaks with *m/z* 416 (M+H+2) and 417 (M+H+3) of hydroxy acid L with MW 413 (**4**; R<sub>1</sub>, R<sub>2</sub>, R<sub>3</sub>=CH<sub>3</sub>) which is a constituent of pamamycin-607 compared with those of non-labeled precursors (Table 2). These results showed that hydroxy acids **3** and **7** were incorporated into **4**, confirming that **1** was biosynthesized from **3** via **7** and **4**.

#### Quantitative Analysis of Hydroxy Acid Intermediates

Hydroxy acids which are precursors of pamamycin in cultured materials of the wild type strain were quantified by LC-ESI-MS analysis when pamamycin production reached to the maximum. The authentic hydroxy acid K (**3**; MW 384; R<sub>1</sub>, R<sub>2</sub>, R<sub>3</sub>=CH<sub>3</sub>), hydroxy acid L (**4**; MW 413; R<sub>1</sub>, R<sub>2</sub>, R<sub>3</sub>=CH<sub>3</sub>) or hydroxy acid S (**5**; MW 230; R<sub>4</sub>=CH<sub>3</sub>, R<sub>5</sub>=H) were obtained from the hydrolysate of the mixture of pamamycin homologues (**1**), and from PM-ketone (**2**) by preparative HPLC, respectively. As a result, the molar ratio of **3**, **4**, **5** and **1** was 5 : 2 : 8 : 200 (Table 3).

## Discussion

Biosynthetic intermediates of pamamycin, hydroxy acids K (**3**), L (**4**), and S (**5**) were obtained from cultured materials of mutants and the wild type strain of *S. alboniger*, but the macrodiolide without nitrogen, PM-ketone (**2**) was not detected. The detected hydroxy acids, **3**, **4** and **5** and de-*N*-methylhydroxy acid L (**7**) were incorporated into pamamycin (**1**), and hydroxy acids **3** and **7** were also converted into hydroxy acid **4**. However, there was no evidence that **2** or de-*N*-methylpamamycin (**6**) were converted to **1**, and **2** was not detected by conversion of hydroxy acids **3** and **5**. These data showed that a nitrogen atom was introduced into the carbonyl group of hydroxy acid K (**3**) to yield hydroxy acid L (**4**), and then hydroxy acid L was condensed with hydroxy acid S (**5**) to produce pamamycin (**1**).

Quantitative analysis of hydroxy acids **3**, **4** and **5** and pamamycin (**1**) suggested that the rate-determining step of pamamycin biosynthesis was a transamination reaction, because accumulation of hydroxy acid L (**4**) was smaller than hydroxy acids K (**3**) or S (**5**), and that of pamamycin was 25~100 times larger than hydroxy acid L. These results showed that the produced hydroxy acid L immediately converted to pamamycin. This regulatory mechanism of pamamycin production is reasonable because introduction of a nitrogen atom has been reported to be essential for aerial mycelium-inducing activity [6, 10].

In some polyketide compounds, the nitrogen-containing unit is incorporated during biosynthesis [12]: the starter unit of tetracyclines is malonamyl-CoA. Rifamycins are biosynthesized from 3-amino-5-hydroxybenzoic acid. Thiazole compound derived from cysteine is the starter unit of epothilones. During biosynthesis of tacrolimus and rapamycin, elongation of the polyketide chain is

terminated by amidation with the nitrogen-containing unit, pipercolic acid. However, only one example reported that a nitrogen atom was introduced into the carbonyl group of a polyketide carbon skeleton, which is the dimethylamino group at C-4 of tetracyclines [13]. Pamamycin is the second example of this type.

The nitrogen atom contributes to expression of various biological activities, which are disclosed by numerous examples of alkaloids and synthetic medicinal and agricultural chemicals. Polyketides are an important group of secondary metabolites including various antibiotics, immunosuppressants, and anticancer agents. Introduction of nitrogen into polyketides will expand potentials of polyketide modifications. There are detailed studies on oxidation of C-6 [14, 15], and chlorination at C-7 [16] in post-PKS tailoring steps of tetracycline biosynthesis. However, there is little information on the transamination reaction [17].

The gene encoding the transamination reaction during pamamycin biosynthesis will serve as a new and powerful tool for combinatorial chemistry in polyketides. Molecular biological research of nonactin which consists of stereoisomeric homologues of hydroxy acid S in pamamycin, was carried by Priestley *et al.* [18] and Shen *et al.* [19]. We postulate that the homologue of *nonS* gene, which encodes the enzyme catalyzing the formation of a tetrahydrofuran ring of the constituent hydroxy acid of nonactin, exists in the pamamycin biosynthetic gene cluster. We are now attempting to clone the gene cluster, and to determine the gene responsible for the transamination reaction.

## Experimental

### General

<sup>13</sup>C-NMR spectra were obtained with a JEOL JNM A-600 spectrometer at 150 MHz in acetone-*d*<sub>6</sub> with an equimolar amount of CF<sub>3</sub>COOD. GC-MS analyses were performed with a JEOL SX-102 A spectrometer. ESI mass spectral data were measured on a JEOLJMS-T100LC AccuTOF mass spectrometer.

### Strain and Media

*S. alboniger* NBRC 12738 and its blocked mutants, strains 188 and 202, which were derived from the wild type strain by *N*-methyl-*N'*-nitro-*N*-nitrosoguanidine (NTG) treatment, were used in this study. These strains were maintained on a Hickey and Tresner's agar (HTA) medium [20].

Medium for pamamycin production and bioconversion experiments contained maltose monohydrate 15.0 g, yeast

extract (Oriental Yeast Co. Ltd.) 3.0 g, polypeptone S (Wako Pure Chemical Industries Ltd.) 10.0 g, NaCl 3.0 g, and CaCO<sub>3</sub> 2.0 g in 1 liter distilled water.

### Mutagenesis

Spores produced by *S. alboniger* NBRC12738 on ISS agar medium (ISP-4) were scraped off, and suspended in 50 mM Tris-HCl (pH 8.0) buffer containing 1% Tween 80. The spore suspension was filtered through No. 2 filter paper and was diluted with the same buffer (1×10<sup>5</sup> spores/ml). The diluted spore suspension was then treated with NTG at 1 mg/ml, and kept standing at 28°C for 1 hour. The treated spore suspension was plated onto HTA medium (2×10<sup>3</sup> spores/9 cm i.d. Petri dish), and incubated at 28°C. After 7 days, colonies in which no or few aerial mycelia formed were transplanted to agar slants. This treatment resulted in a survival rate of *ca.* 1%.

### Selection of Blocked Mutants During Pamamycin Biosynthesis

Response of mutants to pamamycin-607 was examined by the paper disc method. In brief, 4 mutants were radially streaked on HTA medium in a Petri dish (9 cm i.d.), and paper disc containing 10 μg pamamycin mixture (pamamycin-607 content: 15%) was placed in the center. Plates were incubated at 28°C for 7 days, and mutants that restored or stimulated aerial mycelium formation were selected.

Pamamycin production by selected mutants was estimated from the peak area of total ion chromatogram obtained by LC-ESI-MS using pamamycin-607 as standard. Mutants were inoculated into a 500-ml Sakaguchi flask containing 100 ml of the production medium. The flask was placed on a reciprocal shaker (150 strokes/minute) for 6 days at 28°C. Mycelia were collected from cultured broth by filtration, and were treated with acetone and then with EtOAc to obtain the extract. The EtOAc extract was dissolved in MeOH, and was analyzed by LC-ESI-MS as described in the following section.

### LC-ESI-MS Analysis of Pamamycin and Biosynthetic Intermediates

Production and isotopic composition of pamamycin and biosynthetic intermediates were analyzed as follows. LC conditions were: column, Develosil ODS-UG5 (2.0 mm i.d.×150 mm; Nomura Chemical Co. Ltd.); solvent system, 90% aq. MeOH containing 0.2% NH<sub>4</sub>OAc for pamamycin (**1**) and PM-ketone (**2**) or MeOH-0.1% HCOOH (3:2) for hydroxy acids **3**, **4**, **5**, **7**; flow rate, 0.2 ml/minute; column oven temp., 25°C. MS conditions were: detection mode, ESI positive mode; needle voltage, 1500 V; orifice 1

voltage, 49~51 V; ringlens voltage, 9~13 V; desolvation temp., 250°C; orifice temp., 80°C.

### Bioconversion of the Possible Intermediates

Mutants were inoculated into 100 ml of the production medium in a 500-ml Sakaguchi flask, and the flask was placed on a reciprocal shaker (150 strokes/minute) at 28°C for 72 hours. This seed culture (3 ml) was transferred to 500-ml Sakaguchi flasks, each containing 100 ml of the same medium, and the flasks were incubated on a reciprocal shaker under the same conditions. Twelve hours after inoculation, substrate was added to each flask, and cultivation continued for an additional 60 hours. Compound **2**, **5** or **6** was added as MeOH solution (1.0 mg/0.5 ml) and **3**, **4** or **7** was added as the mixture with **5** at the same concentration as mixture. Extraction and analysis of the converted products, pamamycin (**1**) or hydroxy acid L (**4**), were performed as mentioned above. Ratios of molecular ion peak and isotope peaks in the molecular ion cluster were calculated from results of 3~6 runs of LC-ESI-MS analysis.

### Production of Intermediates Produced by Two Mutants and the Wild Type Strain

Pamamycin intermediates were analyzed by LC-ESI-MS using 120-hour cultured material. PM-ketone (**2**) fraction was obtained from mycelia with EtOAc extraction similarly to **1**. The **3**, **4** and **5** homologue fractions were obtained from filtrate and mycelia.

### Preparation of Authentic Compounds for LC-MS

#### Analysis and Substrate for Bioconversion

##### PM-ketone (**2**)

PM-ketone (**2**) was prepared by permanganate oxidation [11] of **1** (composition of homologues, 607 : 621 : 635 : 649 = 1 : 3 : 2 : 1). Potassium permanganate (34 mg, 0.22 mmol) was added to a solution of **1** (4.3 mg, 0.0071 mmol) in *t*-BuOH-H<sub>2</sub>O (1 : 1, 10 ml), and the solution was stirred for 2 hours at room temperature. The reaction mixture was filtered through celite, and this filtrate was treated with EtOAc after being adjusted to pH 8. The EtOAc extract was concentrated *in vacuo* and was applied to a Sep-Pak Plus silica cartridge. Compound **2** was eluted with EtOAc from the cartridge to separate from **1** (2.6 mg; yield, 64%). The structure of **2** was confirmed by disappearance of *N*-CH<sub>3</sub> signals in <sup>1</sup>H- and <sup>13</sup>C-NMR spectra and LC-ESI-MS (*m/z* 579, 593, 607 621 [M+H]<sup>+</sup> and *m/z* 596, 610, 624, 638 [M+NH<sub>4</sub>]<sup>+</sup>). This obtained fraction was used for bioconversion of **2** to **1**.

<sup>13</sup>C-Labeled **2** was prepared from <sup>13</sup>C-labeled **1** in a similar manner as described above. <sup>13</sup>C-labeled **1** was

obtained by a feeding experiment of [2-<sup>13</sup>C] acetate in *S. alboniger* as previously described [8].

##### Hydroxy acid L (**4**) and S (**5**)

Hydroxy acid L (**4**) and S (**5**) were prepared by alkaline hydrolysis of **1** [21]. A solution of **1** (8.8 mg, 0.014 mmol) and potassium hydroxide (3.36 g, 60 mmol) in ethanol-H<sub>2</sub>O (8 : 2, 20 ml) was refluxed for 16 hours. The reaction mixture was cooled, and was extracted three times with CH<sub>2</sub>Cl<sub>2</sub> (50 ml) after being adjusted to pH 6. The aqueous residue was then extracted three times with CH<sub>2</sub>Cl<sub>2</sub> (50 ml) at pH 3. These CH<sub>2</sub>Cl<sub>2</sub> extracts were combined, and concentrated *in vacuo* to obtain **4** and **5**. This crude extract was used for bioconversion of **4** and **5** to **1**.

To confirm the structures **4** and **5**, the crude extract was methylated with diazomethane, and was analyzed by GC-MS (column, DB-1 (0.25 mm i.d.×15 m, film thickness 0.1 μm, J&W Scientific); inlet temp., 250°C; chamber temp., 250°C; column temp.: 50°C for 1 minute, 50°C/minute to 150°C, 16°C/minute to 280°C, 280°C for 5 minutes.) [4, 5]. The extract contained hydroxy acid L (**4**) with MWs 413, 427 and 441, and hydroxy acid S (**5**) with MWs 230, 244 and 258. Proportion of hydroxy acid L with MW 413 (**4**; R<sub>1</sub>, R<sub>2</sub>, R<sub>3</sub>=CH<sub>3</sub>), which is component of pamamycin-607 [6], was 20% of total hydroxy acid L, and that of hydroxy acid S with MW 230 (**5**; R<sub>4</sub>=CH<sub>3</sub>, R<sub>5</sub>=H) was 15% of total hydroxy acid S. This mixture was used for bioconversion of **4** and **5** to **1**.

Hydroxy acids **4** and **5** were separated by preparative TLC (CHCl<sub>3</sub>-MeOH (9 : 1)), and gave **4** (R<sub>f</sub> value, 0~0.20; 3.2 mg; 53%) and **5** (R<sub>f</sub> value, 0.35~0.60; 2.6 mg; 78%). Purified **5** was used for bioconversion to **1** with strain 202. These obtained fractions were used as reference compounds for analyzing **4** or **5** in cultured materials, and in products of bioconversion experiment of **5**. The authentic hydroxy acid L with MW 413 (**4**; R<sub>1</sub>, R<sub>2</sub>, R<sub>3</sub>=CH<sub>3</sub>) or hydroxy acid S with MW 230 (**5**; R<sub>4</sub>=CH<sub>3</sub>, R<sub>5</sub>=H) for quantitative analysis was obtained from each fraction by preparative HPLC (column, Develosil ODS-UG5 (20 mm i.d.×250 mm, Nomura Chemical); solvent, MeOH-0.1% HCOOH (3 : 2 for **4**, 7 : 3 for **5**); flow rate, 10 ml/minute; detection, 220 nm).

<sup>13</sup>C-labeled **4** and **5** were prepared from <sup>13</sup>C-labeled **1** in a similar manner as described above.

##### Hydroxy Acid K (**3**)

Hydroxy acid K (**3**) was prepared by alkaline hydrolysis of **2** (9.0 mg, 0.016 mmol) using a similar method as for the preparation of hydroxy acids **4** and **5** from **1**. The CH<sub>2</sub>Cl<sub>2</sub> extract of the reaction mixture was used for bioconversion of **3** and **5** to **1**.

The product was methylated with diazomethane, and analyzed by GC-MS to confirm the structure of **3** under same conditions used for analysis of hydroxy acids **4** and **5**. Hydroxy acid K (**3**) was obtained as a mixture of homologues with MW 370, 384, 398, and 412; and the content of hydroxy acid K with MW 384 which is the precursor of pamamycin-607 in total hydroxy acid K homologues was about 22% by GC-MS analysis.

The crude extract which contained **3** and **5** was purified by preparative TLC (CHCl<sub>3</sub> - MeOH (8 : 2)), which gave **3** (R<sub>f</sub> value 0.65~0.75, 3.5 mg, 59%). The obtained fraction was used as reference compound for analyzing **3** in cultured materials.

<sup>13</sup>C-Labeled **3** was prepared from <sup>13</sup>C-labeled **2** in a similar manner as described above.

#### De-*N*-methylhydroxy Acid L (**7**)

De-*N*-methylhydroxy acid L (**7**) was prepared by alkaline hydrolysis of de-*N*-methylpamamycin **6** [9, 10], which was isolated from cultured materials of the wild type strain, with a similar manner as for the preparation of hydroxy acids **4** and **5** using 10.0 mg (0.017 mmol) of **6**. The CHCl<sub>2</sub> extract was used for bioconversion of **7** and **5** to **1**.

<sup>13</sup>C-Labeled **7** was prepared from <sup>13</sup>C-labeled **6** in a similar manner as described above. <sup>13</sup>C-labeled **6** was obtained by a feeding experiment of [2-<sup>13</sup>C] acetate in *S. alboniger*.

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